# **Cover Page**

## **Annual Report #2 (2014)**

#### **Crowland Mitigation through Restoration**

## **of the Tamarack Bog, Bath Nature Preserve. Summit County Ohio.**

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# **Appendices**

**Appendix A: Copies of all Field data sheets Appendix B: List of Vouchers and Voucher numbers collected**

**Appendix C: List of Taxa** 

#### **Monitoring Methods and Results**

VIBI Modules- Methods. As in 2013 we used a modified VIBI methodology (Mack 2007) to evaluate wetland quality. Methods follow those in the 2013 report – we repeat those here for convenience. Our modifications largely involve the shape of individual modules to

accommodate the challenging terrain, thick shrub vegetation, and sensitive habitat (especially in the core bog area). We modified the standard 10x10m VIBI module layout as shown in Figure 1. In this modified design we first established a central 25x1m access lane, then sampled 2m on either side of this lane. This design minimized trampling while allowing good access to the 4x25m sampling area.

We established 11 such modules (Figure 2): 3 in the core bog area, 4 adjacent to the wetland edge (near the delineated boundary of the wetland), and 4 that are potential areas of wetland expansion. Our intent was to: 1) use the core modules to evaluate whether the existing bog maintains its status during the restoration. 2) use the wetland Edge modules to evaluate whether conditions at the edge improve (e.g., become more boglike, and experience spread of sphagnum or other bog specialists). 3) use the Expansion modules (which generally had a noticeably peaty soil with a 'bounce', and seemed likely to improve if hydrology was restored) to evaluate wetland quality and the extent of responses to the restoration. We denoted each module in the field with permanent markers, and recorded gps coordinates. We sited modules to include representative habitat of each of the areas listed above.



In each module we used standard VIBI methods to assess presence and percent cover of herbaceous vegetation, along with both percent cover and stem abundance of different size classes of woody plants. We summarized these data using the OEPA's VIBI spreadsheet calculator available online.

VIBI Modules- Results 2014**.** In August 2014 we repeated the 2013 sampling of all 11 100M<sup>2</sup> plots. We identified 140 plant species and another 62 (mostly rare) taxa during our survey, including many peatland specialists (Table 1, Appendix A). We also documented substantial cover by undesirable (e.g., Red Maple, Crabapple), and invasive species (e.g., Buckthorn).



*Figure 2 – Approximate locations of major landscape elements of the tamarack bog restoration. Yellow and orange dashed lines indicate approximate boundaries of major plant communities. Boxes indicate the 11 VIBI modules. The orange boxes are 'core' modules, the yellow boxes are 'edge' modules, and the green boxes are 'enhancement' modules. The 8 green lines indicate vegetation transects. The red T's indicate locations of the 8 tamarack trees.*

*Table 1. Dominant plants (mean relative cover over 5%) from VIBI plots in each wetland area (mean relative cover for each species in parentheses) during 2014.*



In 2014 we did not detect strong changes in the plant community or in of the metrics from the 2013 (year 0) survey. Below are some summaries of our findings for these two initial years of study.

VIBI scores for Core and Edge VIBI areas were high in both years, while the Enhancement areas scored much lower. ANOVA indicates that only the effect of habitat area (core/edge/enhancement) is significant  $(F<sub>2, 16</sub>=$ 84, P<0.001), with no significant differences among years  $(F_{1, 16} = 2.7, P > 0.1)$ , and no interaction (F<sub>2, 16</sub> = 0.8, P>0.4).

FQAI scores were high and showed strong differences among areas that were generally consistent across years, despite a significant interaction of year and area  $(F_{2, 16} = 18,$ P<0.03) that reflects minor improvements in edge plots and declines in enhancement plots. Areas differed significantly  $(F_{2, 16} = 37)$ P<0.001), although years did not  $(F_{1, 16} = 0.4,$  $P>0.5$ ).



Rare species of concern are maintaining themselves, but there is no noticeable numerical growth or spatial spread in 2014

compared to 2013. For example, although the tamarack trees continue to survive and produce cones, we saw no seedling tamarack trees.

The Sphagnum Reach monitoring plots were established in early 2014. Because those results were included in the 2013 report, we do not repeat that information here.

Repeat Photography. In 2014 we established repeat photography stations at each of the 11 VIBI plots. Those photos are included at the end of each of the individual transect sheets (attached). We have also established photo sites at the transect endpoints.

Transects. To evaluate habitat status and future expansion outside of the core bog we resampled 8 transects radiating out from the bog (see Figure 1, green lines). Each transect extended from  $\sim$ 10m inside the delineated wetland boundary to upland habitat (determined by elevation and vegetation). Transects ranged from 40 to 100m in length. NOTE - for this second year of sampling we extended several transects further upland and into the wetland, to better capture potential changes. Every 10m along each transect we scored canopy coverage, hydrology, and soils. We considered each 10m portion of a transect as a 'segment'. We grouped the data across transects using soil description into wetland (25 segments), transition (13 segments), and upland (14 segments). In most cases the transect results match expectations, with clear gradients along

the transition from wetland to upland habitat: litter generally increased, mosses decreased, and canopy increased. We noted minor changes between 2013 and 2014, but because of the expanded coverage of the transects these are not interpretable.



## Hydrological chemistry

Two rounds of water chemistry evaluation have been completed in the tamarack bog, and those data are now being evaluated by UA geologists (Ira Sasowsky and Karyna Mezentseva), and Ohio EPA scientists (Jeff Rizzo and Joe Loucek). They are working to resolve the causes and meaning of an imbalance of cations and anion. That evaluation should be completed by mid to late 2015, when Mezentseva completes her Master's thesis on the hydrology of the bog.

Hydrological monitoring – Methods (From Dr. Ira Sasowsky and Karyna Mezentseva, UA Geology).

Dates: 12/03/13 and 12/05/13.A total of 11 borings were made using Geoprobe direct push method. Initial probing was done using a 2.25" diameter probe that collects a 1.25" diameter sample. Samples were collected from all borings using clear acrylic liners, 48 inches in length. The boring numbers and status are given in the summary table. The liners were cut open in the field for examination. All samples were photographed. The samples were later closed up, and wrapped in Saran wrap for preservation and later testing. It was typical that the first 4' push returned <4' of sample (compression). It was also typical that deeper samples would fill a 4' core with only 3' of push (expansion). The typical sampling core-column was organic matter on top, followed by brown clay, and then gray clay. The gray clay had occasional pebbles in it. A few holes had sand or gravel layers. Many holes were dry. Wells that were completed (i.e. screen and pipe installed) are 1.5" inner diameter white PVC, with pre-packed screens of 5' length. They are installed in a 3.25" diameter pushed hole. Riser pipe is screwed together. Sand was poured in to the annulus after screens were placed, and a weighted line was used to try and allow 2' of sand above top of screen. Annulus was then backfilled to surface with granular bentonite. Date: 03-15-2014. The purpose of field work was to install several hand-drilled wells within the bog boundaries for water level and chemical monitoring. Possible auger well locations were previously selected. Map with wells positions was created in a GIS program Global Mapper. Disto laser distance meter and tape measure along with Brunton bearing (corrected for 8 degree declination) were used to get the approximate position for the new borings. A 4.2 feet long and 0.3 foot width bucket auger, with 3 feet extensions, was used for creating the borings. Total number of installed augured was wells 5. Two wells at different depth (long and shorter) were installed at the spot  $# 7$  and  $8$  in order to calculate hydraulic gradient at those places. Boreholes were made by twisting an auger directly into the peat with subsequent placing of PVC 1" x1'SDR-21 PR 200 PSI pipes (outer and inner diameter s are1.25 and 1.125 inches, respectively). Filters made of dense cotton thread were designed by Tom Quick (Research Associate at the Department of Geosciences, the University of Akron) and attached to the bottoms of the PVC pipes in order to prevent pipes from clogging by sediment. Total length of filter was 13.5". Mud and peat samples were laid out on the yellow cloth, described, collected in Ziploc bags, appropriately labeled. All samples were photographed. Generally, samples made of somewhat muddy layer on the top that gradually changes into wet partially decomposed organic layer sometimes abundant with woody fragments. Consistently graded medium grit sand Arena Mediana and granular bentonite were poured into the annulus to isolate the sampling interval in the wells. Due to the unstable nature of the borehole walls, and the small annular space, it was not possible to quantify the height of sand placed in the screened intervals.



# **Auger wells characteristics**



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## Hydrological monitoring – Results to date.





"-" -not applicable,

a – Only select borings were completed as wells,

b – Measured by GPS placed on the top of casing or stake, 100 point average, do not trust,

c – Elevations derived from Lidar data using Global Mapper,

d – Reported by driller,

e – Measured by tape,

f –Determined by subtracting stick up values from top of the casing wells.

# **Appendix A: Copies of all Field data sheets – see attachment**







**Appendix C.** Taxon List for Tamarack Bog as of 2014. Includes 140 identified species, and 62 taxa not yet confirmed to species. Does not distinguish mosses (approximately 20 species identified in the bog so far).

*Acer rubrum Acer saccharum Agrimonia pubescens Agrimonia striata Alliaria petiolata Alnus incana Alnus serrulata Alnus sp. Amelanchier laevis Amelanchier sp. Amphicarpaea bracteata Arctium sp. Arisaema triphyllum subsp. Triphyllum Aronia melanocarpa Aster lateriflorus Aster puniceus Aster sp. Berberis thunbergii Bidens cernua Bidens sp. Boehmeria cylindrica Botrychium sp. Calamagrostis sp. Caltha palustris Cardamine sp. Carex atlantica subsp. Capillacea Carex bromoides Carex comosa Carex crinita var. crinita Carex cristatella Carex gracillima Carex lacustris Carex leptalea Carex seorsa Carex sp. Carex trisperma Carpinus caroliniana Carya glabra Carya ovata Carya sp. Cephalanthus occidentalis Chelone glabra Cinna arundinacea Circaea lutetiana Clematis virginiana Cornus amomum Cornus florida Cornus racemosa Crataegus alnifolia*

*Crataegus sp Crataegus sp. Cuscuta gronovii Decodon verticillatus Desmodium sp. Dryopteris carthusiana Dryopteris cristata Dryopteris marginalis Epilobium ciliatum Epilobium coloratum Epilobium sp. Erechtites hieracifolia Erigeron annuus Euonymus alatus eupatorium perfoliatum Fern 2 Fern sp. Fraxinus pennsylvanica Fraxinus sp. Fungus sp. Galium aparine Galium asprellum Galium labradoricum Galium sp. Galium triflorum Gaylussacia baccata Geranium maculatum Geum canadense Geum laciniatum Geum sp. Glechoma hederacea Glyceria canadensis Glyceria striata grass sp Grass sp. Hackelia virginiana Herb sp. Holcos lanatus Hydrophyllum virginianum Ilex verticillata Impatiens capensis Impatiens pallida Juglans nigra Juncus effusus Lactuca biennis Larix laricina Leersia oryzoides Leersia virginica Lemna minor Lichen sp. Ligustrum sp.*

*Ligustrum vulgare Lindera benzoin Liverwort sp. Lonicera morrowii Lonicera sp. Loniciera maackii Lycopus sp. Marchantiophyta sp. Mentha arvensis Mimulus ringens Mitchella repens Moss sp. Nyssa sylvatica Onoclea sensibilis Osmunda cinnamomea Osmunda regalis Ostrya virginiana Oxalis sp. Oxalis stricta Parthenocissus quinquefolia Penthorum sedoides persicaria virginiana Phalaris arundinacea Physocarpus opulifolius Pilea pumila Poa pratensis Poa sp. Poaceae sp. Polygonum arifolium Polygonum sagittatum Polygonum virginianum Polystichum braunii Polytrichum Populus deltoides Prunus serotina Prunus sp. Prunus virginiana Pyrus coronaria Pyrus sieboldii Pyrus sp. Quercus alba Quercus rubra quercus sp. Ranunculus sp. Rhamnus alnifolia Rhamnus frangula Ribes americanum Ribes palustre Ribes sp. Rosa multiflora Rosa palustris*

*Rubus allegheniensis Rubus flagelarris Rubus flagellaris Rubus hispidus Rubus occidentalis Rubus setosus Rubus sp. Rumex orbicularis Salix sp. Sambucus canadensis Scutellaria lateriflora Sium suave Solanum dulcamara Solidago canadensis Solidago patula Solidago rugosa Solidago sp.*

*Solidago uliginosa Solidago UNK narrow leaved Sphagnum sp. Spiraea alba Stellaria longifolia Symphyotrichum sp. Symphyotruchum novae-angliae Symplocarpus foetidus Symplyotirchum sp. Taraxacum sp. Thelypteris palustris Tiarella cordifolia Toxicodendron radicans Toxicodendron vernix Triadenum fraseri Typha latifolia Typha sp.*

*Ulmus americana Ulmus rubra Ulmus sp. UNK tree seedling 1 Urtica dioica var dioica Urtica dioica var. procera Vaccinium corymbosum Verbena urticifolia Verbesina alternifolia Viburnum cassinoides Viburnum dentatum Viburnum nudum Viola canadensis Viola sp. Vitis sp. Zelkova serrata*